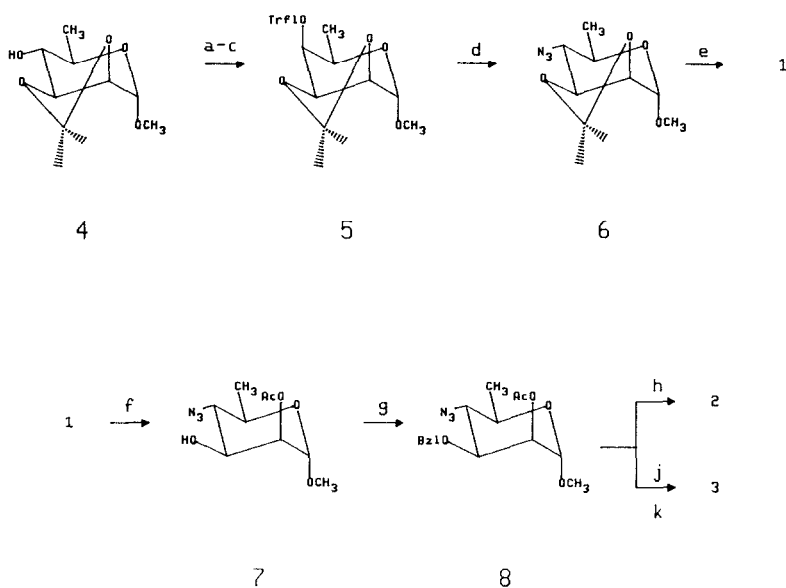


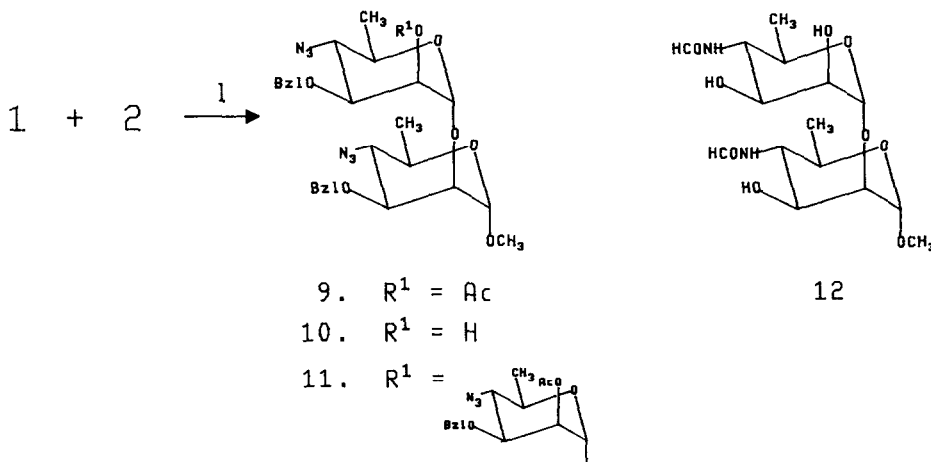
directly from methyl 2,3-*O*-isopropylidene- α -D-mannopyranoside⁶ in 65% yield by iodination at C-6 followed by hydrogenation⁷. A double inversion at C-4 accompanied by introduction of an azide substituent provided the target intermediate (1) via a talopyranoside prepared by pyridinium chlorochromate⁸ or Swern⁹ oxidation, followed by sodium borohydride reduction. Displacement of triflate from the talopyranoside¹⁰ (5) gave the azido derivative (1) 80%, m.p. 79-80° lit.⁴ 81.5-82.5°, $[\alpha]_D^{22} + 127.2^\circ$ (c , 0.98 in CH_2Cl_2) lit.⁴ + 126.9° (in MeOH) after removal of the isopropylidene group from (6). Selective protection for utilization in glycoside synthesis was accomplished by regio-selective acetylation of (1) to give (7) via the 2,3-orthoacetate¹¹. This compound which also serves as a glycosyl acceptor for generation of 1,3 linkages present to some extent in the *Bruceella* M antigen² was benzylated by benzyl trichloroacetimidate¹² under acid catalysis. Transesterification of (8) produced the glycosyl acceptor (2) as an analytically pure syrup, $[\alpha]_D^{22} + 141.0^\circ$ (c , 0.46 in CH_2Cl_2) in 62% overall yield from (4). Acetolysis of glycoside (8) gave quantitatively a mixture of anomeric acetates that were smoothly converted by reaction¹¹ with dichloromethyl methyl ether into the glycosyl chloride (3) 89%, which is suitably protected to direct 1,2-*trans*-glycoside formation and then permits chain extension after glycosylation by removal of the temporary protecting group at O-2. The purity of syrupy glycosyl chloride (3) was established by n.m.r. at 500 MHz.



[a] PCC, powdered molecular sieve, CH_2Cl_2 ; [b] NaBH_4 in MeOH; [c] $(\text{CF}_3\text{SO}_2)_2\text{O}$ (1.17 eq) in CH_2Cl_2 containing pyridine; [d] KN_3 in DMF, 18-crown-6, room temp 2 h; [e] $\text{CF}_3\text{CO}_2\text{H}:\text{H}_2\text{O}$ 9:1, 10 min 0°; [f] (i) $(\text{EtO})_3\text{CMe}$ in DMF/ H^+ ; (ii) 80% aqueous HOAc; [g] $\text{Cl}_3\text{CCNHOCH}_2\text{C}_6\text{H}_4$ in $\text{CCl}_4/\text{cyclohexane}$ $\text{CF}_3\text{SO}_3\text{H}$; [h] NaOMe in MeOH; [j] $\text{Ac}_2\text{O}:\text{HOAc}:\text{H}_2\text{SO}_4$ 100:40:1, room temp 3 h; [k] Cl_2HCOMe , ZnBr_2 in CH_2Cl_2 , room temp 1 h.

SCHEME 1

Glycosyl chloride (3) effectively glycosylated the protected alcohol (2) in a silver triflate promoted reaction to yield the disaccharide (9) 85% $[\alpha]_D^{22} + 99.3^\circ$ (c , 0.94 in CH_2Cl_2), and it was equally efficient in chain extension reactions with the selectively deprotected disaccharide (10) yielding the trisaccharide (11) 83% $[\alpha]_D^{22} + 82.9^\circ$ (c , 1.0 in CH_2Cl_2). The disaccharide was deprotected in two steps by transesterification and hydrogenolysis, and the N-formylated disaccharide (12) $[\alpha]_D^{22} + 39.3^\circ$ (c , 0.5 in MeOH), obtained after a mixed anhydride reaction with its free amino derivative. This disaccharide was an inhibitor of the binding of *Bruceella* A antigen by both infected cow sera and monoclonal antibodies.



[1] $\text{CF}_3\text{SO}_3\text{Ag}$, 4A molecular sieve in CH_2Cl_2 , -30° 1 h and 3 h to room temp.

SCHEME 2

Syntheses of extended oligomers utilizing the strategy outlined here in combination with methodologies for covalent attachment to protein provide ligands and synthetic antigens that are being applied to problems of serodiagnosis and development of superior reagents for bacterial detection. This includes work in progress to characterize the antibody combining site fragment (Fab) derived from monoclonal antibodies that bind the *Bruceella* A antigen and which is the subject of X-ray diffraction studies.

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